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SELECTIVE CLEAVAGE

OF THE

t-BUTYLOXYCARBONYL PROTECTING GROUP

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A disadvantage in the use of the t-butyloxycarbonyl(BOC) group^{1,2} for N-protection in peptide synthesis is that the acidic conditions normally employed for its removal^{3,4,5} also effect the cleavage of the t-butyl ester group. The latter is used mainly to protect the side chain carboxyl groups of aspartic and glutamic acids.^{1,2} Selective protection of side chain carboxyl groups is of increasing importance for the synthesis of glycopeptides in which aspartic and glutamic acids are involved in peptide-sugar links.⁶

We wish to report a method by which the BOC-group can be removed under mild conditions which leave the t-butyl ester group intact. Hofmann <u>et al.</u>⁷ reported that on the column of an aminoacid analyser, BOC-groups were completely hydrolysed but that Y-t-butyl glutamate was only partially hydrolysed. It therefore seemed probable that a similar ion-exchange resin might be used to bring about the selective cleavage of the BOC-group in the presence of the t-butyl ester group.

The removal of the BOC-group from BOC.Phe.Gly.OBu (I, m.p. $100-102^{\circ}$, $[\alpha]_{D}^{25}-8.0^{\circ}$ (C= 1, MeOH), Rf_A 0.84; Found: C,63.7; H,7.6;

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N,7.6. $C_{20}H_{30}N_2O_5$ requires C,63.4; H,7.9; N,7.4%), and from other protected peptides was carried out by the following general method. The compound, in a volume of MeOH/H₂O (1:1) equivalent to about 3/4 of the void volume of a column of Zeo-Carb 225 (H+), was applied to the column which, after 6 hr. at room temperature, was washed with MeOH/H₂O (1:1). This removed starting material. The product, retained on the column, was obtained by elution with MeOH/10% NH₄OH solution (1:1) followed by evaporation of the solvent. In this way, Phe.Gly.OBu (II, Rf_A 0.63, net positive charge: <u>picrate</u> - m.p. 83-85°, $[\alpha]_G^{25}$ +41.7° (C = 1.84, MeOH), Found: C,49.4;H,5.4; N,13.8. $C_{21}H_{25}N_5O_{10}$ requires C,49.7; H,4.9; N,13.8%) was obtained from I in 87% yield. I was completely deblocked by trifluoroacetic acid (TFA) to the free dipeptide (m.p. 71-73°, $[\alpha]_D^{25} +48.8°$ (C = 0.5, H₂O), Rf_A 0.5, net zero charge, Found: C,59.0; H,7.25; N,12.55. $C_{11}H_{14}N_2O_3$ requires C,59.2; H,6.7; N,12.25%).

Coupling of II with BOC.Ala.OH gave BOC.Ala.Phe.Gly.OBu (III, m.p. 110-111°, $[\alpha]_D^{25}$ -30.5°(C = 0.94, CHCl₃), Rf_A 0.89, Rf_B 0.94, Found:C,61.3; H,8.0; N,9.0. $C_{23}H_{34}N_{3}O_{6}$ requires C,61.6; H,7.6; N,9.4%) which was completely deblocked with TFA giving Ala.Phe.Gly.OH (m.p. 160-162°, $[\alpha]_D^{25}$ +45.2° (C = 0.6, MeOH), Rf_A 0.53, net zero charge, Found: C,52.5; H,6,9; N,12.9. $C_{14}H_{19}N_{3}O_{4}$.1 $\frac{1}{2}H_{2}O$ requires C,52.6; H,6.8; N,13.1%. DNP.Ala.Phe.Gly.OH gave m.p. 66-68°, Found: C,52.4; H,4.8. $C_{20}H_{21}N_{5}O_{8}$ requires C,52.4; H,4.6%).

The BOC-group was selectively removed from III giving Ala.Phe.Gly.OBu (IV, Rf_A 0.76, net positive charge: <u>picrate-</u> m.p. 94-96°, $[\alpha]_{G}^{28}$ +90.0° (C = 0.5, MeOH), Found: C,50.4; H,5.0. C₂₄H₃₀N₆O₁₁ requires C,49.85; H,5.2%) in 47% yield. The protected tetrapeptide Z.Gly.Ala.Phe.Gly.OBu (Rf_A 0.88, Rf_B 0.91, $[\alpha]_{D}^{25}$ -7.0°

^{*}Amino-acids, except glycine, were of the L-configuration. Thinlayer chromatography was carried out on MN-silica gel G: Rf_A, pyridine-HAc-BuOH-H₂O (10:5:35:50); Rf_B, BuOH-HAc-H₂O (4:1:5). Paper electrphoresis gave net charges at pH 7.0.

(C = 0.6, CHCl₃), amino-acid ratios in acid hydrolysate $Gly_{1.94}$ Ala_{0.95}Phe_{1.10}) was obtained in 65% yield when IV was coupled with Z.Gly.OH by the mixed anhydride method.²

Selective deprotection of BOC.Ala.Glu $(OBu)_2$ (V, m.p. 87-89°, $[\alpha]_D^{25} - 27.0^\circ$ (C = 1.2, MeOH), Rf_A 0.90, Found: C, 58.8; H, 8.8; N, 6.3. $C_{21}H_{38}N_2O_7$ requires C, 58.6; H, 8.8; N, 6.5%) gave a 63% yield of Ala.Glu. $(OBu)_2$ (VI, Rf_A 0.74, net positive charge: <u>picrate</u> - m.p. 70-72°, Found: C, 46.8; H, 5.9. $C_{22}H_{33}N_5O_{12}$ requires C, 47.4; H, 5.9%) while TFA converted V into the free dipeptide Ala. Glu. $(OH)_2$ (m.p. 92-98°, $[\alpha]_D^{24} - 40.0^\circ$ (C = 0.5, MeOH), Rf_A 0.11, net negative charge, Found: C, 40.9; H, 5.3; N, 11.6. $C_8H_{14}N_2O_5.H_2O$ requires C, 40.6; H, 5.9; N, 11.8%. DNP-Ala.Glu. $(OH)_2$ had m.p. 75-77°, Rf_A0.36, Found: C, 43.8; H, 4.2. $C_{14}H_{16}N_4O_9$ requires C, 43.7; H, 4.7%).

From VI was prepared BOC.Phe.Ala.Glu.(OBu), (VII, m.p. $76-78^{\circ}$, $[\alpha]_{D}^{25} - 31.3^{\circ}$ (C = 1.5, MeOH), Rf_A 0.93, Rf_B 0.95, Found: C, 61.7; H, 8.1; N, 7.6. C₃₀H₄₇N₃O₈ requires C, 62.3; H, 8.1; N, 7.2%), which was completely deblocked with TFA giving the free tripeptide Phe.Ala.Glu.(OH)₂ (m.p. 153-156°, $[\alpha]_D^{25} - 27.0°$ (C = 1.0, MeOH), Rf_A 0.54, net negative charge, Found: C, 53.9; H, 5.9. $C_{17}H_{23}N_{3}O_{6} \frac{1}{2}H_{2}O$ requires C, 54.5; H, 6.4%). Selective removal of the BOC group from VII gave the partially protected tripeptide Phe. Ala.Glu.(OBu)₂ (VIII, $[\alpha]_{D}^{25} - 15.0^{\circ}$ (C = 0.6, benzene), Rf_B 0.54, net positive charge: picrate - m.p. 110-111°, Found: C, 51.5; H, 6.1; N, 11.6. C₃₁H₄₂N₆O₁₃ requires C, 51.5; H, 5.8; N, 11.7%) in 72% yield, and VIII was coupled with Z-(nitro)Arg.Ala.OH⁸ by the mixed anhydride method. This gave the protected pentapeptide Z-(nitro) Arg.Ala.Phe.Ala.Glu.(OBu)₂, $[\alpha]_{D}^{25}$ - 15.0° (C = 0.5, MeOH), Rf_B 0.95, paper chromatography - Rf_A 0.93, Rf_B 0.96. On acid hydrolysis the pentapeptide gave amino-acid ratios: Nitroarg1.06Ala2.03Phe0.95 Glu0.97 (nitroarginine analysed as ornithine and arginine).

The method therefore promises to extend the usefulness of

the BOC and t-butyl ester groups for glycopeptide synthesis and for organic chemistry in general.

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REFERENCES

- E. Schroder and K. Lubke, "The Peptides", Vol. 1, Academic Press, New York, 1965.
- K. Hofmann and P.G. Katsoyannis in "The Proteins", ed. H. Neurath, 2nd. Ed., Vol.1, p.53, Academic Press, New York, 1963.
- ³ G.W.Anderson and A.C.McGregor, J.Am.Chem.Soc. <u>79</u>, 6180, (1957).
- ⁴ F.C.McKay and N.F.Albertson, <u>J.Am.Chem.Soc., 79</u>, 4686, (1957).
- 5 R.Schwyzer and H. Kappeler, <u>Helv.Chim.Acta.</u> <u>44</u>, 1991 (1961); H.Kappeler and R.Schwyzer, <u>ibid</u>, <u>43</u>, 1453 (1960).
- 6 "Glycoproteins", ed. A. Gottschalk, Vol. 5, Elsevier, Amsterdam, 1966, p. 273.
- 7 K.Hofmann, R. Schmiechen, R.D.Wells, Y.Wolman and N.Yanihara, J.Am.Chem.Soc., 87, 611 (1965).
- 8 K.Hofmann, W.D.Peckham and A.Rheiner, <u>J.Am.Chem.Soc.</u>, <u>78</u>, 238 (1956).

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